SERVICES INCORPORATED

/ 7451 Third Street / Detroit, MI 48202 /

Bernie Bowers
Wyandotte Public Schools
639 Oak Street
Wyandotte, MI 48192

Date Reported: 03/02/21
EKS Job: 14852

RE: Roosevelt High School
540 Eureka, Wyandotte, MI 48192

Overview

On February 26, 2021 EKS Services Incorporated (EKS) was retained by Bernie Bowers of Wyandotte Public Schools to perform mold air and tape lift sampling at the location listed above. The air samples were collected and taken to an accredited laboratory and analyzed pursuant to IMS Laboratory Analytical Method: 2.2 (method for analyzing spore trap). The lift samples were collected and taken to an accredited laboratory and analyzed pursuant to IMS Analytical Method: 2.6.1 (method for analyzing abundant organisms tape lift).

Following this letter, please find the Chain of Custody Record that identifies the sample locations and the Laboratory Report for the samples. Mold air samples were collected indoors in the 1st, 2nd, and 3rd floor hallways, the annex hall, the music wing, and the cafeteria. EKS also collected a sample "outdoors", outside of the music wing, to serve as a comparison. EKS included a field blank as a quality control measure of the sampling media and the laboratory process. EKS collected a single tape lift sample from the register grill at the north entrance of B123.

Results/Findings

The table below identifies the locations of the samples collected along with the results.

Mold Air Samples

Sample Number	Location of Sample	Total Count	Spores/M^3
1	Field Blank	No Spores Detected	No Spores Detected
2	3rd Floor Hallway By A308	3 – Pen/Asp group 1 – Ascospores 1 – Basidiospores 5 – Total	160 – Pen/Asp group 53 – Ascospores 53 – Basidiospores 266 – Total
3	2nd Floor Hallway By A207	7 – Basidiospores 5 – Pen/Asp group 1 – Chaetomium 1 – Cladosporium 14 – Total	373 – Basidiospores 267 – Pen/Asp group 53 – Chaetomium 53 – Cladosporium 746 – Total
4	1st Floor Hallway By A107	No Spores Detected	No Spores Detected
5	Annex Hall Across from B110	3 – Cladosporium 1 – Basidiospores 1 – Pen/Asp group 5 – Total	160 – Cladosporium 53 – Basidiospores 53 – Pen/Asp group 266 – Total
6	Music Wing by C137	1 – Pen/Asp group	53 – Pen/Asp group

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Sample Number	Location of Sample	Total Count	Spores/M^3
7	Exterior By Music Wing Entrance	2 – Pen/Asp group 1 – Basidiospores 3 – Total	107 – Pen/Asp group 53 – Basidiospores 160 – Total
8	North side of Cafeteria	No Spores Detected	No Spores Detected

The table below identifies the location of the tape sample collected along with the result.

Tape Lift Sample Result

Sample Number	Location of Sample	Sample Identification	Spores/M^3
1.1	Register Grill at North Entrance B123	No Fungal Spores Observed	

For "clean buildings" total indoor spore concentrations are usually less than 2,000 Spores/M³ with *Aspergillus/Penicillium* ideally comprising less than half the total of indoor spore concentrations, or 1,000 Spores/M³. In this case, the samples taken, at the time of sampling, were indicative of a "clean building". The indoor samples were comparable with the outdoor sample, which had 107 Spores/M³ of *Aspergillus/Penicillium* and a total spore concentration of 160 Spores/M³. No sample had *Stachybotrys*, or the mold known as "black mold", on it.

Abatement Considerations

EKS recognizes that currently no Occupational Safety and Health Administration (OSHA) or Environmental Protection Agency (EPA) standards for mold remediation exist. Solely based on the air and tape lift sample results, EKS did not find a need for any concern with regards to mold indoors at the time of sampling.

Additional Information

Mold spores can easily become airborne and can be found almost anywhere. Because they are so small, mold spores may invade the protective mechanisms of the nose and upper respiratory tract. Mold spores can float through a building forming new colonies, wherever they land.

For fungi/mold to grow, multiple elements are needed such as the presence of spores, water availability, food sources, temperature, presence of competing organisms, light, and pH. By changing or eliminating one or more of these items one can reduce or even prevent the fungi/mold from growing. The conditions for causing mold such as water leaks, condensation, infiltration, or flooding should be corrected to prevent mold from growing. Some recommendations to eliminate or head off the potential for water and moisture build up are as follows:

- Implementing the use of air conditioners or dehumidifiers during humid months.
- Making sure the structure has adequate ventilation, including exhaust fans in the bathrooms that exhaust to the outside.
- Using mold inhibitors in paints.
- Cleaning bathrooms with mold killing products or a 10% bleach and water solution.*
- Never installing carpet in bathrooms.
- Removing and replacing flooded carpets.
- Removing and replacing any water damaged porous building materials.

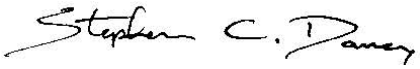
* Care should be used when using any type of chemical products to eliminate mold.

Even though serious health problems can occur, allergy-like symptoms have been reported among individuals that are exposed to mold. These can include nasal and sinus congestion, coughing, breathing

difficulties, sore throat, skin and eye irritation, upper respiratory infections, trigger asthma, respiratory illness, shortness of breath, and tightness of the chest.

The results given in this report apply only to the areas tested at the time of testing. This report should not be reproduced, except in full, without permission of EKS Services Incorporated. It has been a pleasure assisting you. If you have any questions, please contact me at (313) 963-1433.

Sincerely,



Stephen Dancy
Project Manager

3130 Old Farm Lane, Suite 1
Commerce Twp., MI 48390

877-665-3373

Laboratory Report

Prepared Exclusively For:

EKS Services INC.
7451 Third Street
Detroit, MI 48202
(313) 963-1433
info@eksservices.com

Project: 14852 **Lab # E156784**

Report Date: 03/01/2021 **Sampled: 02/26/2021** **Received: 02/26/2021** **Analyzed: 03/01/2021**

Table of Contents

1 - Description of Testing Methods

Air Samples - Spore Trap Sampling: Nonviable spore trap cassette sampling impacts nonviable particles directly into a spore trap cassette at a predetermined flow rate and time. After the sampling period, the cassette is analyzed at IMS Laboratory through direct microscopic examination by a qualified mycologist. Because the analysis does not include culturing the fungi, the results include both viable and non-viable spores. Spore trap samples identify particles, pollen and fungal elements. High particulates in the air can result in underestimation of spore concentrations. This collection methodology is biased toward larger spore sizes. Because some fungal spores cannot be distinguished by direct microscopic examination (e.g. *Penicillium* and *Aspergillus*), these organisms are grouped into larger categories. Examples include the Pen/Asp group, Basidiospores, and Ascospores. Results from this methodology are reported as spores per cubic meter.

Topical Samples - Lift Tape Sampling: Lift tape samples are nonviable surface samples collected by carefully pressing cellophane tape onto a surface to lift surface contaminants and then placing the tape onto a laboratory microscope slide. The lift tape is analyzed by a mycologist at IMS Laboratory through direct microscopic examination. The mycologist then reports the mold spores observed on the surface of a material. This testing methodology is used to identify the types and relative proportions of mold on a surface.

2 - Laboratory Results

Location: Field Blank

Sample # E156784 - 1

Medium Type: AllergencoD
Serial # 1
Exposure: 0.00 l/min. for 0.00 min.
Reporting Limit: 1 Spore/cu. m

Sample Identification	Raw Count	Spores/cu. m	Percent(%)
- Fungi -			
No Trace or Fungal Spores Observed	0	0	N/A%

Background Item	Level
Dust / Debris	Very Low
Opaque Particles	Very Low

Location: 3rd Floor Hall by A308

Sample # E156784 - 2

Medium Type: AllergencoD
Serial # 2
Exposure: 15.00 l/min. for 5.00 min.
Reporting Limit: 53 Spores/cu. m

Sample Identification	Raw Count	Spores/cu. m	Percent(%)
- Fungi -			
Pen/Asp group	3	160	60.15%
Ascospores	1	53	19.92%
Basidiospores	1	53	19.92%
Total Fungi	5	266	100.00%
- Other -			
Pollen	1	53	100.00%

Background Item	Level
Dust / Debris	Medium
Opaque Particles	Low

Location: 2nd Floor Hall by A207

Sample # E156784 - 3

Medium Type: AllergencoD
Serial # 3
Exposure: 15.00 l/min. for 5.00 min.
Reporting Limit: 53 Spores/cu. m

Sample Identification	Raw Count	Spores/cu. m	Percent(%)
- Fungi -			
Basidiospores	7	373	50.00%
Pen/Asp group	5	267	35.79%
Chaetomium	1	53	7.10%
Cladosporium	1	53	7.10%
Total Fungi	14	746	100.00%

Background Item	Level
Dust / Debris	Medium
Opaque Particles	Low

Location: 1st Floor Hall by A107

Sample # E156784 - 4

Medium Type: AllergencoD
Serial # 4
Exposure: 15.00 l/min. for 5.00 min.
Reporting Limit: 53 Spores/cu. m

Sample Identification	Raw Count	Spores/cu. m	Percent(%)
- Fungi -			
No Fungal Spores Observed	0	0	N/A%

Background Item	Level
Dust / Debris	Low
Opaque Particles	Very Low

Location: Annex Hall Across B110

Sample # E156784 - 5

Medium Type: AllergencoD
Serial # 5
Exposure: 15.00 l/min. for 5.00 min.
Reporting Limit: 53 Spores/cu. m

Sample Identification	Raw Count	Spores/cu. m	Percent(%)
- Fungi -			
Cladosporium	3	160	60.15%
Basidiospores	1	53	19.92%
Pen/Asp group	1	53	19.92%
Total Fungi	5	266	100.00%

Background Item	Level
Dust / Debris	Medium
Opaque Particles	Low

Location: Music Wing by C137

Sample # E156784 - 6

Medium Type: AllergencoD
Serial # 6
Exposure: 15.00 l/min. for 5.00 min.
Reporting Limit: 53 Spores/cu. m

Sample Identification	Raw Count	Spores/cu. m	Percent(%)
- Fungi -			
Pen/Asp group	1	53	100.00%

Background Item	Level
Dust / Debris	Low
Opaque Particles	Low

Location: Exterior Music Wing Ent.

Sample # E156784 - 7

Medium Type: AllergencoD
Serial # 7
Exposure: 15.00 l/min. for 5.00 min.
Reporting Limit: 53 Spores/cu. m

Sample Identification	Raw Count	Spores/cu. m	Percent(%)
- Fungi -			
Pen/Asp group	2	107	66.88%
Basidiospores	1	53	33.13%
Total Fungi	3	160	100.00%

- Sample data continued on next page -

Background Item	Level
Dust / Debris	Low
Opaque Particles	Low

Location: Cafeteria N. Side

Sample # E156784 - 8

Medium Type: AllergencoD
Serial # 8
Exposure: 15.00 l/min. for 5.00 min.
Reporting Limit: 53 Spores/cu. m

Sample Identification	Raw Count	Spores/cu. m	Percent(%)
- Fungi -			
No Fungal Spores Observed	0	0	N/A%

Background Item	Level
Dust / Debris	Very Low
Opaque Particles	Very Low

Location: Room B123 Register Grill at N. Entrance

Sample # E156784 - 9

Medium Type: Tape Lift
Serial # 1.1

Sample Identification	Prevalence
- Fungi -	
No Fungal Spores Observed	

Background Item	Level
Dust / Debris	Very Low
Opaque Particles	Very Low

Analytic Methods and Formulas:

Calculated results may include one more significant figure than is mathematically justified in order to accommodate the client's needs.
IMS Analytical Method: 2.6.1 (method for analyzing abundant organisms tape lift)
IMS Laboratory Analytical Method: 2.2 (method for analyzing spore trap)
Spores per cubic meter is determined by: Total Spore Count x 4000 / (sampling rate x sampling time)

Note that this report may use mold-specific units of measure, such as Spores/cu. m and CFU/cu. m, for Sample Identifications which are not mold. Examples include pollen, fabric and fiberglass fibers, insect particles, and ash. In this context, "CFU" and "Spore" refer to individual pieces of the identified material.

IMS Laboratory, LLC is accredited through the AIHA-LAP, LLC and participates in Environmental Microbiology Proficiency Testing, EMPAT #172958. Data is provided in compliance with AIHA-LAP, LLC policy modules and ISO/IEC 17025:2017 guidelines.

Kathryn C. Langley 03/01/2021

Kathryn C. Langley, Laboratory Manager

3 - Spore Trap Comparison Chart

SAMPLING LOCATIONS

1: Field Blank
2: 3rd Floor Hall by A308
3: 2nd Floor Hall by A207
4: 1st Floor Hall by A107
5: Annex Hall Across B110
6: Music Wing by C137

Spores per Cubic Meter

Mold Name \ Location #	1	2	3	4	5	6
Alternaria						
Arthrinium						
Ascospores		53				
Basidiospores		53	373		53	
Bipolaris / Drechslera group						
Chaetomium			53			
Cladosporium			53		160	
Curvularia						
Erysiphe/Oidium						
Fusarium						
Ganoderma						
Mitospores						
Pen/Asp group		160	267		53	53
Pithomyces						
Polythrincium						
Rust						
Smuts/Periconia/Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Unknown Fungi						
FUNGAL TOTAL	**0**	**266**	**746**	**0**	**266**	**53**
Pollen		53				

Please refer to the Laboratory Results section for additional details.

SAMPLING LOCATIONS

7: Exterior Music Wing Ent.
8: Cafeteria N. Side

Spores per Cubic Meter

Mold Name \ Location #	7	8
Alternaria		
Arthrinium		
Ascospores		
Basidiospores	53	
Bipolaris / Drechslera group		
Chaetomium		
Cladosporium		
Curvularia		
Erysiphe/Oidium		
Fusarium		
Ganoderma		
Mitospores		
Pen/Asp group	107	
Pithomyces		
Polythrincium		
Rust		
Smuts/Periconia/Myxomycetes		
Stachybotrys		
Stemphylium		
Torula		
Unknown Fungi		
FUNGAL TOTAL	**160**	**0**
Pollen		

Please refer to the Laboratory Results section for additional details.

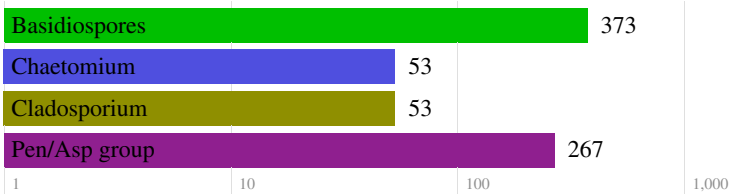
4 - Sample Comparison Graph

Spore Trap Samples - Spores per Cubic Meter

Field Blank
NO FUNGI FOUND

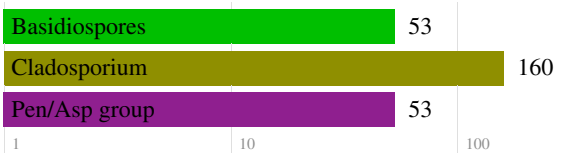
3rd Floor Hall by A308

2nd Floor Hall by A207



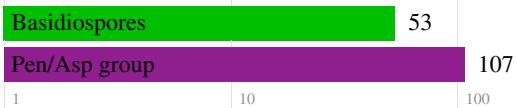
1st Floor Hall by A107
NO FUNGI FOUND

Annex Hall Across B110



Music Wing by C137

Exterior Music Wing Ent.



Spore Trap Samples - Spores per Cubic Meter

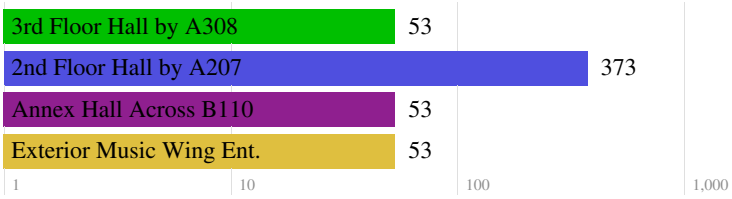
Cafeteria N. Side
NO FUNGI FOUND

5 - Background Comparison Graph

Spore Trap Samples - Spores per Cubic Meter

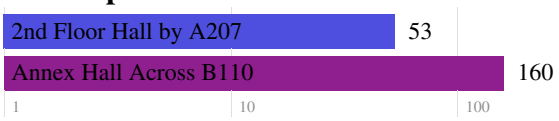
Ascospores

Basidiospores

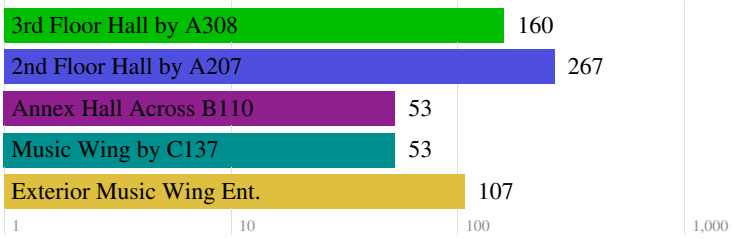


Chaetomium

Cladosporium



Pen/Asp group



Pollen

6 - Understanding Laboratory Results

Laboratory findings must only be considered as part of an overall mold investigation. The interpretation of the findings must only be made by a qualified individual after reviewing all relevant data. Visual information and environmental conditions measured during the site assessment are crucial to any final interpretation of the results. A very good reference book which covers sampling and data interpretation has been published by The American Conference of Governmental and Industrial Hygienists and is entitled *Bioaerosols: Assessment and Control*, 1999.

Numerical guidelines cannot be used as the primary determinant as to whether a mold problem may exist. Concentrations of mold in the air will vary depending on weather conditions, building air flow, time of day and time of year. Comparisons between indoor and outdoor mold levels, types of mold found, visual information and environmental conditions are more important in interpreting results than reliance on specific numeric thresholds.

In *Indoor Air Quality in Office Buildings: A Technical Guide*, Health Canada, Revised 1995 (Pages 49-50), Health Canada set forth guidelines which can be used to better understand air testing results. The guidelines included these general principles. Significant numbers of certain pathogenic fungi should not be present in indoor air (e.g. *Aspergillus fumigatus*, *Histoplasma*, and *Cryptcoccus*). Bird or bat droppings in air intakes, ducts or rooms should be assumed to contain these pathogens. The persistent presence of significant numbers of toxigenic fungi (e.g. *Stachybotrys atra*, toxigenic *Aspergillus*, *Penicillium* and *Fusarium* species) indicate that further investigation and action should be taken. The confirmed presence of one or more fungal species occurring as a significant percentage of a sample in indoor air samples and not similarly present in concurrent outdoor samples is evidence of a fungal amplifier. The "normal" air mycoflora is qualitatively similar and quantitatively lower than that of outdoor air. The significant presence of fungi in humidifiers and diffuser ducts and on moldy ceiling tiles and other surfaces requires investigation and remedial action regardless of the airborne mold concentrations.

Generally, mold spores are present everywhere. As a general rule, "normal" air mycoflora is qualitatively similar and quantitatively lower than that of outdoor air. When the converse is true, it is likely that an indoor source of mold may exist. However, even this most basic rule may produce misleading results. Airborne mold spore levels vary widely due to factors such as weather conditions and activity levels. For example, in a "normal" home, indoor mold spore levels may be elevated above outdoor spore levels after vacuuming (when airborne indoor levels could be unusually high) or after a heavy snow (when outdoor levels could be unusually low).

Surface Sampling primarily identifies the types and relative proportions of mold on a surface. Viable surface sampling will identify living mold, while nonviable surface sampling will identify all mold (but cannot distinguish between living or dead mold). Surface sampling may confirm that a substance is mold or identify the types of mold present on the surface. Because mold is everywhere, there is a high probability that a surface sample from a "clean" surface will still identify mold on that surface.

There are currently no state or federal standards or guidelines regarding results of fungal samples. There are no levels, which are typical or permissible. There are no recommended exposure limits, no permissible exposure limits, no threshold limit values and no short term exposure limits.

These guidelines are not intended, nor should they be used, for health evaluation purposes or to evaluate the safety of an occupied space. A physician should be consulted regarding health and/or safety questions.

7 - Sample Identification Definitions

Ascospores

A large group of spores that are very ubiquitous in nature. They are frequently found in the air after a rain. Most ascospores are plant pathogens; a small portion have been known to cause infection in humans but are identified separately.

Found in these Sample Locations: (2) 3rd Floor Hall by A308

Basidiospores

A large group of spores that are very ubiquitous in nature. They are released from mushrooms, shelf fungi, puffballs, and a variety of other macro fungi. Basidiospores may be allergenic to those with seasonal allergies.

Found in these Sample Locations: (2) 3rd Floor Hall by A308 (3) 2nd Floor Hall by A207 (5) Annex Hall Across B110 (7) Exterior Music Wing Ent.

Chaetomium

A type of ascospore commonly isolated from soil. It is found on a variety of substrates including decomposing plant material and wood, dung, straw, and damp or water-damaged cellulose (e.g. paper on drywall). As a moisture-indicator fungi, Chaetomium only grows when the substrate has a current or previous severe moisture problem. There are over 100 documented species of Chaetomium, several of which are reported to be toxigenic; if not speciated, the genus Chaetomium should be assumed to be toxigenic. It has been known to cause systemic, cerebral, cutaneous, subcutaneous, and pulmonary infections, though usually only in the immunocompromised.

Found in these Sample Locations: (3) 2nd Floor Hall by A207

Cladosporium

One of the most commonly identified outdoor fungi. It is often found indoors in numbers less than outdoors. Cladosporium is also found on decaying plants and food, straw, paint, and textiles. It is generally regarded to be allergenic and can be a cause of extrinsic asthma (immediate type hypersensitivity: Type I). Cladosporium has been reported in cases of skin lesions, keratitis, onychomycosis, sinusitis, and pulmonary infections.

Found in these Sample Locations: (3) 2nd Floor Hall by A207 (5) Annex Hall Across B110

No Fungal Spores Observed

Analyst did not observe matter which could be identified as fungal spores.

Found in these Sample Locations: (4) 1st Floor Hall by A107 (8) Cafeteria N. Side (9) Room B123 Register Grill at N. Entrance

No Trace or Fungal Spores Observed

No visible trace or spores were observed in the sampling area.

Found in these Sample Locations: (1) Field Blank

Pen/Asp group

The spores of the genera *Penicillium*, *Aspergillus*, and *Trichoderma* are quite similar when viewed under a microscope and are grouped together under the heading Pen/Asp. *Penicillium* species are among the most common fungi found in indoor environments, particularly basements. Certain species may cause infections of the eye, external ear, respiratory system, and urinary tract. Some species of *Aspergillus* are parasitic on insects, plants, and animals including humans. All *Aspergillus* species are allergenic. Various species can cause extrinsic asthma, pulmonary emphysema, opportunistic infections of the ears and eyes, and severe pulmonary infections. Some species of *Penicillium*, *Aspergillus*, and *Trichoderma* produce mycotoxins which may be associated with diseases in humans and animals. Several toxins are considered potential human carcinogens. The genus *Trichoderma* has been reported to cause infections in immunocompromised individuals, patients undergoing dialysis, and individuals with chronic kidney failure or chronic lung disease.

Found in these Sample Locations: (2) 3rd Floor Hall by A308 (3) 2nd Floor Hall by A207 (5) Annex Hall Across B110 (6) Music Wing by C137 (7) Exterior Music Wing Ent.

Pollen

Pollen are coarse to fine particles/grains produced by various trees, weeds, and grasses. For individuals with seasonal allergies, pollen is often the causative agent.

Found in these Sample Locations: (2) 3rd Floor Hall by A308

8 - Warranties, Legal Disclaimers, and Limitations

IMS's scope of accreditation through the AIHA-LAP, LLC is for the following FoT(s) / Method(s): Fungal Air - Direct Examination (SOP 2.2 and 2.3); Fungal Bulk - Direct Examination (SOP 2.6); and Fungal Surface - Direct Examination (SOP 2.1).

The study and understanding of molds is a progressing science. Because different methods of sampling, collection and analysis exist within the indoor air quality industry, different inspectors or analysts may not always agree on the mold concentrations present in a given environment. Additionally, the airborne levels of mold change frequently and by large amounts due to many factors including activity levels, weather, air exchange rates (indoors), and disturbance of growth sites. It is possible for report interpretations and ranges of accuracy to vary since comprehensive, generally accepted industry standards do not currently exist for indoor air quality inspections of mold in residential indoor environments. This report is intended to provide an analysis based upon samples taken at the site at the time of the inspection. Mold levels can and do change rapidly, especially if home building materials or contents remain wet for more than 24 hours, or if they are wet frequently. This report is not intended to provide medical or healthcare advice. All allergy or medical-related questions and concerns, including health concerns relating to possible mold exposure, should be directed to a qualified physician. If this report indicates indoor mold levels that are higher than in typical indoor living spaces relative to the outdoor environment, or indicates any findings that are of concern to you, further evaluation by a trained mold professional or a Certified Industrial Hygienist (CIH) may be advisable.

Results pertain only to the samples tested, as received by IMS. Unless otherwise noted in the body of this report, the condition of samples upon receipt was acceptable. Blank samples are reported in the same manner as all other samples. The results are not corrected for contamination.

This report is generated by IMS at the request of, and for the exclusive use of, the IMS client named on this report. Project Name, Project Number, Sampling Date, Sampling Locations and Exposure times and rates have been provided to IMS by the client, and may affect the validity of the results. The analysis of the test samples is performed by IMS. This report applies only to the samples taken at the time, place and location referenced in the report and received by IMS, and to the property and weather conditions existing at that time only. Please be aware, however, that property conditions, inspection findings and laboratory results can and do change over time relative to the original sampling due to changing conditions, the normal fluctuation of airborne mold, and many other factors. IMS does not furnish, and has no responsibility for, the inspector or inspection service that performs the inspection or collects the test samples. It is the responsibility of the end-user of this report to select a properly trained professional to conduct the

inspection and collect appropriate samples for analysis and interpretation. Neither IMS, nor its affiliates, subsidiaries, suppliers, employees, agents, contractors and attorneys ("IMS related party") are able to make and do not make any determinations as to the safety or health condition of a property in this report. The client and client's customer are solely responsible for the use of, and any determinations made from, this report, and no IMS related party shall have any liability with respect to decisions or recommendations made or actions taken by either the client or the client's customer based on the report.

Samples analyzed by IMS are disposed the day that they are analyzed. Storage may be available for a fee with written request at the time the samples are submitted for analysis.

IMS hereby expressly disclaims any and all representations and warranties of any kind or nature, whether express, implied or statutory, related to the testing services or this report including, but not limited to, damages for loss of profit or goodwill regardless of the negligence (either sole or concurrent) of IMS and whether IMS has been informed of the possibility of such damages, arising out of or in connection with IMS's services or the delivery, use, reliance upon or interpretation of test results by client or any third party. In no event will IMS be liable for any special, indirect, incidental, punitive, or consequential damages of any kind regardless of the form of action whether in contract, tort (including negligence), strict product liability or otherwise, arising from or related to the testing services or this report.

IMS accepts no legal responsibility for the purposes for which the client uses the test results. IMS will not be held responsible for the improper selection of sampling devices even if we supply the device to the user. The user of the sampling device has the sole responsibility to select the proper sampler and sampling conditions to insure that a valid sample is taken for analysis. Additionally, neither this report nor IMS makes any express or implied warranty or guarantee regarding the inspection or sampling done by the inspector, the qualifications, training or sampling methodology used by the inspector performing the sampling and inspection reported herein, or the accuracy of any information provided to IMS serving as a basis for this report. The total liability of IMS related to or arising from this report to a client or any third party, whether under contract law, tort law, warranty or otherwise, shall be limited to direct damages not to exceed the fees actually received by IMS from the client for the report. The invalidity or unenforceability, in whole or in part, of any provision, term or condition herein shall not invalidate or otherwise affect the enforceability of the remainder of these provisions, terms and conditions. Client shall indemnify IMS and its officers, directors and employees and hold each of them harmless for any liability, expense or cost, including reasonable attorney's fees, incurred by reason of any third party claim in connection with IMS's services, the test result data or its use by client.

- End of Lab Report Number E156784 -

CHAIN OF CUSTODY RECORD

E156784

EKS SERVICES INCORPORATED
7451 THIRD STREET
DETROIT, MI 48202
(313)963-1433 PHONE

CLIENT: EKS SERVICES	PROJECT MANAGER: STEVE DANCY
ADDRESS: SEE ABOVE	PHONE: SEE ABOVE

JOB #: 14852

REQUIRED TURNAROUND TIME
PLEASE CIRCLE ONE

1 HR | 3 HR | 4 HR | 6 HR
24 HR | 48 HR | (3 DAY) | 5 DAY

ATTENTION LABORATORY: IF THE CIRCLED TURNAROUND TIME IS NOT AVAILABLE PLEASE CALL AND ADVISE WHAT TURNAROUND IS AVAILABLE. THANK YOU.

MATERIAL DESCRIPTION	SAMPLE NUMBER	ASBESTOS PLM	ASBESTOS TEM / AHERA / LEVEL II / NIOSH 7402	LEAD AIR	MOLD / BIOAEROSOL/AIR / BULK / TAPE / VACUUM	VOLUME	LEAD PAINT
FIELD BLANK	1					0	
3RD FLOOR HALL BY A308	2					75	
2nd FLOOR HALL BY A207	3					75	
1ST FLOOR HALL BY A107	4					75	
ANNEX HALL ACROSS B110	5					75	
MUSIC WING BY C137	6					75	
EXTERIOR MUSIC WING ENT.	7					75	
CAFETERIA N. SIDE	8					75	
ROOM B123 REGISTER	1.1				TAPE		
GRILL AT N. ENTRANCE							

RELINQUISHED BY: Michael B. Pace	DATE: 2-26-21	TIME: 1045	RECEIVED BY:	DATE: RECEIVED FEB 26 2021	TIME:
RELINQUISHED BY:	DATE:	TIME:	RECEIVED BY:	DATE:	TIME:

REVISED 04/29/14